

Didemnidines A and B, Indole Spermidine Alkaloids from the New Zealand Ascidian *Didemnum* sp.

Rhys Finlayson,[†] A. Norrie Pearce,[†] Michael J. Page,[‡] Marcel Kaiser,[§] Marie-Lise Bourguet-Kondracki,[⊥] Jacque L. Harper,^{||} Victoria L. Webb,[∇] and Brent R. Copp^{*,†}

[†]Department of Chemistry, University of Auckland, Private Bag 92019, Auckland, New Zealand

[‡]National Institute of Water & Atmospheric Research (NIWA) Ltd, PO Box 893, Nelson, New Zealand

[§]Swiss Tropical and Public Health Institute, Socinstrasse 57, PO Box CH-4002, Basel, Switzerland, and University of Basel, CH-4003 Basel, Switzerland

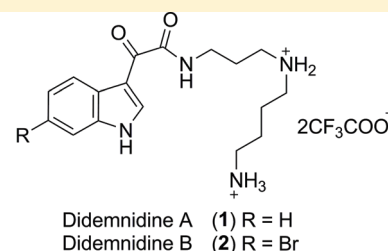
[⊥]Laboratoire des Molécules de Communication et Adaptation des Micro-organismes, FRE 3206 CNRS, Muséum National d'Histoire Naturelle, 57 Rue Cuvier (C.P. 54), 75005 Paris, France

^{||}Malaghan Institute of Medical Research, PO Box 7060, Wellington South, New Zealand

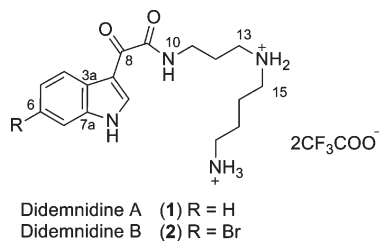
[∇]NIWA, Private Bag 14-901, Kilbirnie, Wellington, New Zealand

S Supporting Information

ABSTRACT: Two new indole spermidine alkaloids, didemnidines A (1) and B (2), have been isolated from the New Zealand ascidian *Didemnum* sp. The structures of the metabolites, determined by analysis of 2D NMR spectra and confirmed via synthesis, embody an indole-3-glyoxylamide moiety linked to the N¹ position of spermidine, the latter motif being particularly rare among marine natural products. Didemnidine B and a synthetic precursor exhibited mild in vitro growth inhibition of *Plasmodium falciparum* with IC₅₀'s of 15 and 8.4 μM, respectively.



Marine natural product chemistry of ascidians of the family Didemnidae is dominated by amino acid-derived secondary metabolites,^{1,2} which include cyclic peptides, pyridoacridine, β-carboline, lamellarin, 3,4-substituted maleimide, and polysulfide alkaloids. In many cases, these natural products exhibit biological activities with either human therapeutic or ecological relevance.² In the context of our search for potential anti-inflammatory lead molecules from New Zealand marine organisms,³ we have investigated a mildly bioactive extract prepared from a *Didemnum* sp. ascidian collected at Tiwai Point, Southland, New Zealand. In addition to the known maleimides didemnimides A–D,⁴ two new indole-3-glyoxylamidospersmidine analogues, didemnidines A (1) and B (2), were isolated. The structures of both natural products were confirmed by synthesis, and they and two synthetic intermediates were tested for bioactivity in a range of anti-inflammatory, antitumor, and neglected disease whole parasite assays.



Extraction of the freeze-dried ascidian sample with MeOH, followed by fractionation utilizing C₈, C₁₈, and Sephadex LH-20

column chromatography, led to the purification of didemnimides A–D⁴ and didemnidines A (1) and B (2). Our initial attempts to dereplicate the didemnimides via HPLC-DAD, ESIMS, and NMR were hampered somewhat by the fact that we had isolated the alkaloids as their TFA salts, whereas the original isolation report⁴ presented data for the alkaloids as free bases. Considerable differences were observed for the ¹H NMR resonances assigned to imidazole proton H-16 (Δδ_{salt-free base} +1.0 to +1.4) and benzenoid proton H-4 (Δδ_{salt-free base} –0.3 to –0.4) between the salt and free base forms. Rigorous 2D-NMR spectroscopic assignments however determined the identities of the alkaloids (see Supporting Information for details).

Didemnidine A (1), isolated as a brown oil, displayed a pseudomolecular ion at *m/z* 317.1980 [M + H]⁺ in the (+)-HRESI mass spectrum, establishing a molecular formula of C₁₇H₂₄N₄O₂. An absorption maximum at 328 nm in the UV spectrum suggested the presence of an indole moiety. The ¹H NMR spectrum of 1 (Table 1) exhibited resonances attributable to seven exchangeable protons (δ_H 12.29, 8.90, 8.54 (2H), 7.81 (3H)), five aromatic methines, and 14 alkyl protons. The ¹³C NMR spectrum contained signals for all 17 carbons required by the molecular formula. Analysis of COSY NMR correlations established the presence of three separate proton spin systems: four protons (δ_H 8.23, 7.55, 7.27, and 7.25) were part of a 1,2-disubstituted benzene ring, two coupled proton resonances (δ_H

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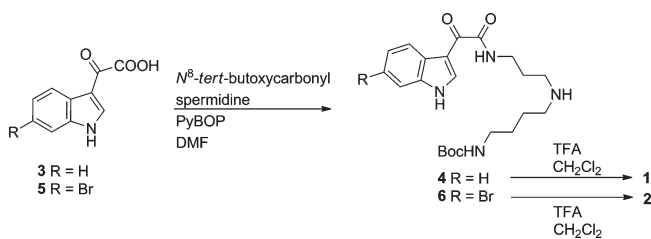
Table 1. ^1H (600 MHz), ^{13}C (150 MHz), and HMBC NMR Data for Didemnidines A (1) and B (2)^a

position	didemnidine A (1)			didemnidine B (2)		
	δ_{C}	δ_{H} (J in Hz)	HMBC ^b	δ_{C}	δ_{H} (J in Hz)	HMBC ^b
1		12.29, br s			12.39, s	
2	138.4	8.77, d (3.1)	3, 3a, 7a, 8	139.2	8.78, s	3, 3a, 7a
3	112.0			112.0		
3a	126.1			125.2		
4	121.1	8.23, m	3, 3a, 7a	122.8	8.15, d (8.4)	3, 3a, 6, 7a
5	122.5	7.25, m	3a, 7	125.4	7.41, dd (8.4, 1.5)	3a, 7
6	123.4	7.27, m	7, 7a	115.9		
7	112.5	7.55, m	3a, 7a	115.3	7.75, d (1.5)	5, 6, 7a
7a	136.2			137.1		
8	181.6			181.7		
9	163.7			163.3		
10		8.90, t (6.2)	9, 11		8.92, t (6.2)	9, 11
11	35.7	3.30 (obscured) ^c	12, 13	35.8	3.30 (obscured) ^c	12, 13
12	25.6	1.86, m	11, 13	25.6	1.85, m	11, 13
13	44.6	2.94, br s		44.6	2.94, br m	12
14		8.54, 2H, br s			8.49, 2H, br s	
15	46.0	2.94, br s		46.0	2.94, br m	16, 17, 18
16	22.5	1.60, br m	15, 17, 18	22.5	1.59, br m	15, 17
17	24.1	1.56, br m	15, 16, 18	24.1	1.59, br m	15, 16, 18
18	38.1	2.80, m	16, 17	38.1	2.81, br s	
19		7.81, 3H, br s			7.79, 3H, br s	

^a Spectra recorded in DMSO-*d*₆ at 27 °C. ^b HMBC correlations, optimized for 8.3 Hz, are reported from the proton resonance to the indicated carbon resonance(s). ^c Resonance obscured by H₂O peak. The corresponding resonances for synthetic 1 and 2 were observed at δ_{H} 3.30 (dt, *J* = 6.2, 6.2 Hz, H₂-11). See Supporting Information.

12.29, 8.77) indicated the presence of a pyrrole ring, and a 20-proton spin system was consistent with the presence of an alkyl polyamine residue. Correlations observed in HSQC and HMBC NMR experiments established that the first two proton spin systems comprised a 3-substituted indole moiety. Among these, correlations observed between resonances assigned to H-4 (δ_{H} 8.23) and δ_{C} 112.0 (C-3) and δ_{C} 136.2 (C-7a), between H-7 (δ_{H} 7.55) and δ_{C} 126.1 (C-3a), and between H-2 (δ_{H} 8.77) and C-3, C-3a, and C-7a were decisive in establishing the indole ring system. In addition, a correlation observed between H-2 and δ_{C} 181.6 (C-8) in the HMBC spectrum established the presence of a carbonyl group at the indole 3-position. Analysis of chemical shifts and correlations observed in COSY, HSQC, and HMBC NMR experiments for the alkyl polyamine residue determined it to represent an *N*¹-amidospemidine fragment (C-9 to (+)NH₃-19).⁵ Interpretation of the COSY spectrum led to one contiguous polyamine substructure extending from an exchangeable proton resonance at δ_{H} 8.90 (t, *J* = 6.2 Hz, NH-10) to δ_{H} 3.30 (obscured by solvent, H₂-11) through H₂-12 (δ_{H} 1.86) and H₂-13 (δ_{H} 2.94) to an exchangeable protonated secondary amine resonance (δ_{H} 8.54, 2H), which in turn correlated to carbon-bound methylene δ_{H} 2.94 (H₂-15) and sequentially through δ_{H} 1.60 (H₂-16), δ_{H} 1.56 (H₂-17), and δ_{H} 2.80 (H₂-18) before terminating at a protonated primary amine resonance (δ_{H} 7.81, br s, H₃-19). HMBC correlations observed between H₂-11 (δ_{H} 3.30) and NH-10 (δ_{H} 8.90) and an amide ^{13}C resonance (δ_{C} 163.7, C-9) established the polyamine to be an *N*¹-amido-substituted spermidine unit. Between them, the 3-carbonyl-substituted indole and *N*¹-amidospemidine fragments accounted for all atoms required by the molecular formula, with a possible connection between C-8 and C-9 of 1. However, no long-range

Scheme 1. Synthesis of Didemnidines A (1) and B (2)



^1H - ^{13}C NMR correlation was observed between NH-10 and C-8 to directly support the presence of this linkage. Comparison of ^{13}C chemical shift data observed for the C-2 to C-9 fragment of 1 with structurally related indole-3-glyoxylamides leptoclidinamine A⁶ and polyandrocarpamide A⁷ showed excellent agreement, allowing the confident assignment of the structure of didemnidine A as shown. The structure of the natural product was confirmed by synthesis, whereby coupling of 2-(indol-3-yl)glyoxylic acid (3)⁸ with *N*⁸-*tert*-butoxycarbonylspermidine^{9,10} in the presence of PyBOP afforded *N*¹-(indolyl-3-glyoxamido)-*N*⁸-*tert*-butoxycarbonylspermidine (4) in 47% yield (Scheme 1). Subsequent removal of the *tert*-butoxycarbonyl protecting group with TFA/CH₂Cl₂ yielded 1 as the *N*¹⁴, *N*¹⁹-difluoroacetate salt. Direct comparison of ^1H and ^{13}C NMR data acquired for this synthetic product with those observed for the natural product (see Supporting Information) confirmed the structure of 1.

Didemnidine B (2) was isolated as a brown oil. A molecular formula of C₁₇H₂₃BrN₄O₂ was assigned to 2 on the basis of the

Table 2. Antiparasitic and Cytotoxic Activities of 1, 2, 4, and 6

	IC ₅₀ (μM)				
	<i>T. b. rhod.</i> ^a	<i>T. cruzi</i> ^b	<i>L. don.</i> ^c	<i>P. falc.</i> K1 ^d	L6 ^e
1	59	130	>180	41	24
2	44	82	>160	15	25
4	34	88	>190	32	73
6	9.9	28	>160	8.4	25
melarsoprol ^f	0.010				
benznidazole ^f		1.35			
miltefosine ^f			0.52		
chloroquine ^f				0.20	
podophyllotoxin ^f					0.01

^a *Trypanosoma brucei rhodesiense*, STIB 900 strain, trypomastigotes stage. ^b *Trypanosoma cruzi*, Tulahuen C4 strain, amastigotes stage. ^c *Leishmania donovani*, MHOM-ET-67/L82 strain, amastigote/axenic stage. ^d *Plasmodium falciparum*, K1 strain, IEF stage. ^e L6 rat skeletal myoblast cell line. ^f Melarsoprol, benznidazole, miltefosine, chloroquine, and podophyllotoxin were used as positive controls.

pseudomolecular ion cluster observed in the (+)-HRESI mass spectrum. Close inspection of ¹H and ¹³C NMR spectra and analysis of the full suite of 2D-NMR data (Table 1) established that the only differences between the natural products 1 and 2 were localized in the indole ring. Analysis of the COSY NMR spectrum indicated the presence of a 1,2,4-trisubstituted benzene ring, consistent with 2 being either a C-5- or C-6-brominated analogue of 1. Consideration of ¹H NMR chemical shifts¹¹ as well as HMBC correlations observed between the proton resonance δ_H 8.15 (d, *J* = 8.4 Hz, H-4) and the quaternary aromatic carbon resonances δ_C 112.0 (C-3), δ_C 115.9 (C-6), and δ_C 137.1 (C-7a) established that didemnidine B was the 6-bromo analogue of didemnidine A. The synthesis of didemnidine B was achieved in a similar manner to that used for didemnidine A, starting this time with 2-(6-bromoindol-3-yl)-glyoxylic acid (5)^{8,12} (Scheme 1). Deprotection of the *tert*-butoxycarbonyl-protected intermediate 6 yielded didemnidine B as the bistrifluoroacetate salt (see Supporting Information).

The occurrence of indole-3-glyoxylamides in marine organisms has been reported five times to date, with simple analogues being reported from the sponge *Spongosorites* sp., coscinamides A–C isolated from the sponge *Coscinosidera* sp., and the ascidian-derived metabolites polyandrocarpamides A–C (*Polyandrocarpa* sp.), 6-bromo-5-hydroxyindolyl-3-glyoxylate (*Syncarpa oviformis*), and leptoclinidamines A and B from *Leptoclinides durus*.⁶ Of the limited number of spermidine derivatives reported from invertebrate and microbial marine organisms,¹³ 1 and 2 join the ranks of squalamine¹⁴ and analogues¹⁵ as being the only examples of mono-*N*¹-substituted analogues.

Didemnidines A (1) and B (2), the *tert*-butoxycarbonyl-protected reaction intermediates 4 and 6, and didemnidines A–D were all found to be inactive when evaluated for the ability to inhibit the superoxide respiratory burst in a model of gout and were not considered cytotoxic in testing in the Developmental Therapeutics Program, NCI. The smaller compound set of 1, 2, 4, and 6 was found to be inactive in screening as phospholipase A₂ and farnesyltransferase enzyme inhibitors. Evaluation against the neglected disease parasite targets *Trypanosoma brucei rhodesiense*, *T. cruzi*, *Leishmania donovani*, and *Plasmodium falciparum* (K1 chloroquine-resistant strain) indicated didemnidine B (2) to

be mildly active toward the malaria parasite (IC₅₀ 15 μM), with slightly enhanced activity being detected for the more lipophilic *tert*-butoxycarbonyl-protected derivative 6 toward *P. falciparum* (IC₅₀ 8.4 μM) and *Trypanosoma brucei rhodesiense* (IC₅₀ 9.9 μM) (Table 2). Both 2 and 6 displayed moderate cytotoxicity toward the nonmalignant L6 cell line, establishing limited selectivity toward the parasites. The ease of synthesis of 6 using a route amenable to the preparation of analogues provides the opportunity to more fully explore the structure–activity relationship of diacylated spermidines as antimalarial and antitrypanosomal agents.

EXPERIMENTAL SECTION

General Experimental Procedures. Ultraviolet–visible spectra were run as MeOH solutions on a UV-2102 PC Shimadzu UV–vis scanning spectrophotometer. Infrared spectra were recorded using a Perkin-Elmer spectrum One Fourier-transform IR spectrometer as a dry film. NMR spectra were recorded on either a Bruker Avance DRX-600 spectrometer operating at 600 MHz for ¹H nuclei and 150 MHz for ¹³C nuclei, a Bruker Avance DRX-400 spectrometer operating at 400 MHz for ¹H nuclei and 100 MHz for ¹³C nuclei, or a Bruker Avance DRX-300 spectrometer operating at 300 MHz for ¹H nuclei and 75 MHz for ¹³C nuclei. Proto-deutero solvent signals were used as internal references (DMSO-*d*₆: δ_H 2.50, δ_C 39.43; CD₃OD: δ_H 3.30, δ_C 49.05; CDCl₃: δ_H 7.25, δ_C 77.0). Standard Bruker pulse sequences were utilized. HRMS data were acquired on either a VG-7070 or a Bruker micrOTOF Q II mass spectrometer. Flash column chromatography was performed using reversed-phase Merck Lichroprep RP-8 or RP-18 40–63 μm, and size exclusion chromatography on Pharmacia Biotech Sephadex LH-20. Analytical reversed-phase HPLC was run on a Waters 600 HPLC photodiode array system using an Alltech C₈ column (3 μm Econosphere Rocket, 33 × 7 mm) and eluting with a linear gradient of H₂O (0.05% TFA) to MeCN over 13.5 min at 2 mL/min and monitoring at 330 nm.

Animal Material. The ascidian was collected by scuba at a depth of 7 m from wooden piles at Tiwai Wharf, Tiwai Point, Southland, New Zealand (46°35.52' S, 168°21.14' E) on August 20, 2007, and kept frozen until used. A voucher specimen of the orange, encrusting irregular shaped ascidian *Didemnum* sp. is held at the National Institute for Water and Atmospheric Research, Private Bag 14-901, Kilbirnie, Wellington, New Zealand, as MNP9167. A taxonomic description and in situ underwater photograph of the organism are included in the Supporting Information.

Isolation and Purification. A single specimen of freeze-dried ascidian material (29.3 g) was extracted with MeOH (7 × 200 mL). The solvent was filtered and then removed under reduced pressure to give a dark brown extract (3.29 g). This extract was subjected to combinations of reversed-phase C₈ flash, C₁₈ flash (MeOH, H₂O–TFA (0.05%)), and Sephadex LH-20 (MeOH, 0.05% TFA) chromatography, yielding didemnimide A (6.2 mg, 0.02% dry weight, *t*_R 4.77 min), didemnimide B (4.8 mg, 0.02% dry weight, *t*_R 4.75 min), didemnimide C (8.3 mg, 0.03% dry weight, *t*_R 4.82 min), didemnimide D (12.1 mg, 0.04% dry weight, *t*_R 4.74 min), didemnidine B (2) (2.0 mg, 0.007% dry weight, *t*_R 5.45 min), and didemnidine A (1) (1.1 mg, 0.004% dry weight, *t*_R 5.65 min).

Didemnidine A (1): brown oil; UV (MeOH) λ_{max} (log ε) 255 (3.49), 266 (3.42), 275 sh (3.35), 328 (3.04) nm; ¹H and ¹³C NMR (see Table 1); (+)-ESIMS *m/z* 317 [M + H]⁺; (+)-HRESIMS *m/z* 317.1980 [M + H]⁺ (calcd for C₁₇H₂₅N₄O₂, 317.1972).

Didemnidine B (2): brown oil; UV (MeOH) λ_{max} (log ε) 259 (4.05), 276 (4.04), 325 (3.90) nm; ¹H and ¹³C NMR (see Table 1); (+)-ESIMS *m/z* 395 ([M + H]⁺, 50%), 397 ([M + H]⁺, 50%); (+)-HRESIMS *m/z*

z 395.1075 [M+H]⁺ (calcd for C₁₇H₂₄⁷⁹BrN₄O₂, 395.1077), 397.1054 (calcd for C₁₇H₂₄⁸¹BrN₄O₂, 397.1057).

*N*¹-(Indolyl-3-glyoxamido)-*N*⁸-*tert*-butoxycarbonylspermidine (**4**). *N*⁸-*tert*-Butoxycarbonyl spermidine^{9,10} (18.1 mg, 73.4 μmol), 2-(indol-3-yl)glyoxylic acid (**3**)⁸ (11.6 mg, 61.2 μmol), and PyBOP (35.0 mg, 67.1 μmol) were dissolved in dry DMF (10 mL), triethylamine (34.1 μL) was added, and the reaction was left to stir at room temperature for 24 h. The crude reaction mixture was subjected to combinations of C₈, C₁₈, and Sephadex LH-20 (MeOH, H₂O–TFA (0.05%)) column chromatography to afford *N*¹-(indolyl-3-glyoxamido)-*N*⁸-*tert*-butoxycarbonylspermidine trifluoroacetate salt (**4**) (15.3 mg, 47%) as a pale yellow oil: IR (neat) ν_{\max} 3389, 1677, 1437, 1204, 1136 cm⁻¹; ¹H NMR (CD₃OD, 400 MHz) δ 8.79 (1H, s, H-2), 8.29 (1H, m, H-4), 7.47 (1H, m, H-7), 7.26 (2H, m, H-5 and H-6), 3.46 (2H, m, H₂-11), 3.05 (6H, m, H₂-13, H₂-15, H₂-18), 1.98 (2H, m, H₂-12), 1.71 (2H, m, H₂-16), 1.56 (2H, m, H₂-17), 1.41 (9H, s, OBU^t); ¹³C NMR (CD₃OD, 100 MHz) δ 181.5 (C-8), 166.5 (C-9), 158.7 (C-1'), 139.6 (C-2), 138.0 (C-7a), 128.0 (C-3a), 125.0 (C-6), 124.0 (C-5), 123.0 (C-4), 114.0 (C-3), 113.2 (C-7), 80.2 (C-2'), 48.4 (C-15, obscured by solvent), 46.6 (C-13), 40.5 (C-18), 36.9 (C-11), 28.8 (OBU^t), 28.1 (C-17), 27.6 (C-12), 24.6 (C-16); (+)-ESIMS *m/z* 417 [M + H]⁺; (+)-HRESIMS *m/z* 417.2509 [M + H]⁺ (calcd for C₂₂H₃₃N₄O₄, 417.2496).

Didemnidine A (**1**). To a solution of **4** (10.1 mg, 19.1 μmol) in CH₂Cl₂ (20 mL) was added TFA (10 mL). The mixture was stirred at room temperature for 70 min, solvent was removed under reduced pressure, and the crude product was purified by Sephadex LH20 column chromatography (MeOH + 0.05% TFA) to yield *didemnidine A* (**1**) as the bistrifluoroacetate salt (7.2 mg, 70%, yellow oil): IR (neat) ν_{\max} 3351, 1672, 1633, 1495, 1436, 1200, 1141 cm⁻¹; UV, ¹H and ¹³C NMR data were identical to those observed for the isolated natural product; (+)-ESIMS *m/z* 317 [M + H]⁺; (+)-HRESIMS *m/z* 317.1973 [M + H]⁺ (calcd for C₁₇H₂₅N₄O₂, 317.1972).

2-(6-Bromoindol-3-yl)glyoxylic acid (**5**). 6-Bromoindole¹² (0.535 g, 2.72 mmol) in a 100 mL two-necked round-bottomed flask was dissolved in anhydrous ether (30 mL). A solution of oxalyl chloride (2.72 mmol) in anhydrous ether (10 mL) was added dropwise to the rapidly stirring indole solution at 0 °C under a nitrogen atmosphere over a period of 10 min. The mixture was stirred for an additional 10 min, and an orange precipitate formed. Saturated NaHCO₃ (10 mL) was added dropwise to the solution and the mixture then heated to reflux for 30 min. Upon cooling, the reaction mixture was extracted with CH₂Cl₂ (3 × 10 mL) and then acidified with 10% HCl(aq) (10 mL) to yield **5** as a yellow precipitate, which was then filtered and dried (0.650 g, 89%): mp 233–234 °C (dec); IR (neat) ν_{\max} 3203, 1712, 1634, 1512, 1410, 1269 cm⁻¹; ¹H NMR (DMSO-*d*₆, 400 MHz) δ 12.63 (1H, s, NH), 8.38 (1H, d, *J* = 3.1 Hz, H-2), 8.05 (1H, d, *J* = 8.7 Hz, H-4), 7.76 (1H, d, *J* = 1.4 Hz, H-7), 7.37 (1H, dd, *J* = 8.7, 1.4 Hz, H-5); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 181.1 (C-8), 165.2 (C-9), 138.9 (C-2), 137.8 (C-7a), 125.9 (C-5), 124.8 (C-3a), 123.0 (C-4), 116.5 (C-6), 115.8 (C-7), 112.5 (C-3); (–)-ESIMS *m/z* 266 [M – H][–] (50%), 268 [M – H][–] (50%); (–)-HRESIMS *m/z* 265.9460 [M – H][–] (calcd for C₁₀H₅⁷⁹BrNO₃, 265.9458), 267.9443 (calcd for C₁₀H₅⁸¹BrNO₃, 267.9438).

*N*¹-(6-Bromoindolyl-3-glyoxamido)-*N*⁸-*tert*-butoxycarbonylspermidine (**6**). *N*⁸-*tert*-Butoxy carbonylspermidine^{9,10} (22.0 mg, 89.7 μmol), 2-(6-bromoindol-3-yl)glyoxylic acid (**5**) (20.1 mg, 75.3 μmol), and PyBOP (40.0 mg, 76.8 μmol) were dissolved in dry DMF (10 mL), triethylamine (40 μL) was added, and the reaction was left to stir at room temperature for 24 h. The crude reaction mixture was subjected to combinations of C₈ flash, C₁₈ flash, and Sephadex LH-20 (MeOH, H₂O–TFA (0.05%)) column chromatography to afford *N*¹-(6-bromoindolyl-3-glyoxamido)-*N*⁸-*tert*-butoxycarbonylspermidine trifluoroacetate (**6**) (18.2 mg, 40%) as a pale yellow oil: IR (neat) ν_{\max} 3388, 1675, 1460, 1205, 1131 cm⁻¹; ¹H NMR (CD₃OD, 400 MHz) δ 8.79 (1H, s, H-2), 8.20 (1H, d, *J* = 8.3 Hz, H-4), 7.66 (1H, d, *J* = 1.6 Hz, H-7),

7.36 (1H, dd, *J* = 8.3, 1.6 Hz, H-5), 3.45 (2H, m, H₂-11), 3.05 (6H, m, H₂-13, H₂-15, H₂-18), 1.98 (2H, m, H₂-12), 1.70 (2H, m, H₂-16), 1.55 (2H, m, H₂-17), 1.41 (9H, s, OBU^t); ¹³C NMR (CD₃OD, 100 MHz) δ 182.1 (C-8), 166.0 (C-9), 158.7 (C-1'), 140.1 (C-2), 138.8 (C-7a), 127.0 (C-5), 126.9 (C-3a), 124.4 (C-4), 118.1 (C-6), 116.2 (C-7), 114.0 (C-3), 80.1 (C-2'), 48.8 (C-15), 46.5 (C-13), 40.5 (C-18), 36.9 (C-11), 28.7 (OBU^t), 28.1 (C-17), 27.5 (C-12), 24.6 (C-16); (+)-ESIMS *m/z* 495 [M + H]⁺ (50%), 497 [M + H]⁺ (50%); (+)-HRESIMS *m/z* 495.1597 [M + H]⁺ (calcd for C₂₂H₃₂⁷⁹BrN₄O₄, 495.1601), 497.1579 (calcd for C₂₂H₃₂⁸¹BrN₄O₄, 497.1582).

Didemnidine B (**2**). To a solution of **6** (9.8 mg, 16.1 μmol) in CH₂Cl₂ (20 mL) was added TFA (10 mL). The mixture was stirred at room temperature for 70 min, solvent was removed under reduced pressure, and the crude product was purified by Sephadex LH20 column chromatography (MeOH + 0.05% TFA) to yield *didemnidine B* (**2**) as the bistrifluoroacetate salt (7.0 mg, 70%, yellow oil): IR (neat) ν_{\max} 3254, 2954, 1672, 1605, 1504, 1125 cm⁻¹; UV, ¹H and ¹³C NMR data were identical to those observed for the isolated natural product; (+)-HRESIMS *m/z* 395.1069 [M + H]⁺ (calcd for C₁₇H₂₄⁷⁹BrN₄O₂, 395.1077), 397.1049 (calcd for C₁₇H₂₄⁸¹BrN₄O₂, 397.1057).

Biological Assays. Details of the neutrophil anti-inflammatory assay,¹⁶ PLA₂ and farnesyltransferase assays,¹⁷ and whole organism parasite assay protocols¹⁸ have been reported elsewhere.

■ ASSOCIATED CONTENT

S Supporting Information. Taxonomic description and color in situ photo of the *Didemnum* sp. ascidian, assigned ¹H and ¹³C NMR data (DMSO-*d*₆) for didemnidines A–D as their trifluoroacetate salts, ¹H, ¹³C, COSY, HSQC, and HMBC NMR spectra (DMSO-*d*₆) for didemnidines A (**1**) and B (**2**), ¹H NMR spectra of synthetic intermediates **4** and **6**, and ¹H and ¹³C NMR spectra for synthetic **1** and **2**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

*Tel: +64 9 373 7599, ext 88284. Fax: +64 9 373 7422. E-mail: b.copp@auckland.ac.nz.

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